

In vivo bioluminescence imaging has become a cornerstone technology for preclinical molecular imaging. This imaging method is based on light-emitting enzymes, luciferases, which require specific substrates for light production. When linked to a specific biological process in an animal model of human biology or disease, the enzyme-substrate interactions become biological indicators that can be studied noninvasively in living animals [1]. Aminoluciferin (*aLuc*) is american firefly (*Photinus pyralis*) luciferin with its 6-position hydroxyl group substitued with an amino group. This modification allows *aLuc* to form amide bond with a peptide, while at the same time retaining the transport and bioluminescent properties of luciferin, resulting in a molecule called peptide-aminoluciferin. Many, particially protected peptide-aminoluciferin (*e.g. Z-Asp-Glu-Val-Asp-aLuc, Z-Leu-Glu-His-Asp-aLuc, Suc-Leu-Val-Tyr-aLuc*) are good substrates for bioluminescence assays, for example in the detection of protease activity. Proteases represent important pharmaceutical targets because of their involvement in numerous disease processes [2]. The above mentioned conjugates generally offer significant advantages, such as increased sensitivity, ease of use, and high throughput screening capacity. Luciferase-based assays are typically 10- to 100-fold more sensitive than comparable fluorescent assays (Rhodamine 110, AMC and AFC) [3].

Light emission of the peptide-aminoluciferin



The synthesis of different type peptide-6-amino-D-luciferin conjugates and their precursors have been published [2] and some of them are commercially available. However, because of their high price the *in vivo* application of these conjugates is limited. To solve this problem we successfully worked out a new, easier and more convenient and economical method for the preparing these derivatives starting from 2-chloro-benzothiazole. Moreover this products have excellent purity (>99%) and adequate yield (82-93%). ATP Mg²⁺ References Biochemistry, 2006, 45, 11103-11112.; [2] J. of Biomol. Sc., 2005, 10(2), 137-148.
BioTechniques, 2011, 51(2), 105-110. Synthesis cc. H₂SO₄ / KNO₃ aq. NH₄Cl / Fe-powder ¹H-NMR spectra of 2-chloro-6-nitro-benzothiazole EtOAc / reflux 0-10°C KCN / KI / DMF / 120° C / 10 h Hal KCN / 18-crown- 6 / DMF / 120 ° C / 8 h Hal 7.4 7.2 ppm Fmoc-AA / isobutyl-chloroformate D-Cys / MeOH RP-HPLC-chromatogram of the purified c-Asp(O'Bu)-6-amino-2-cyano-benzothia Fmoc-AA / TCFH / DCM / rt / overnight rt / 25 min t_R= 25.50 mir Fmoc Fmoc-AA / TFFH / DCM / rt / overnight 2-CI-Trt resin DIPEA / DMF / rt / 3 h н t₀= 19.20 mir NH--Fmoc 20% piperidine / DMF 20% piperidine / DMF Z-AA / DCC / HOBt / DMF Fmoc-AA / HI DCC / HOBt / DMF peptide Fmoc Z-Asp-Glu-Val-Asp-6-amino-luciferin ESI-mass spectra of the 870.3 = [M + H]* 50% TFA / 40% AcN / 10% H 20 45 min / rt -peptide z peptid **Biological tests** Caspase-3/7 activity (24 h) Acknowledgement In vivo: In vitro: ed SCID mice Primary rat retina Luminescenc 500000 ted with A375luc me cell culture + toxir (thapsigargin)

Z-DEVD-aLuc

300000

control

ed with tox

measurement

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